

SHORT REPORT

Open Access

Tungsten-enhanced growth of *Methanosphaera stadtmanae*

Bédis Dridi¹, Saber Khelaifia¹, Marie-Laure Fardeau², Bernard Ollivier² and Michel Drancourt^{1*}

Abstract

Background: The methanogenic *Archaea Methanosphaera stadtmanae* has been detected in the human gut microbiota by both culture and culture-independent methods. Its growth reaches an exponential phase after 5 to 7-day culture in medium 322 (10% vol). Our recent successful isolation of *Methanomassiliicoccus luminyensis*, a tungstate-selenite-requiring *Archaea* sharing similar metabolism characteristics with *M. stadtmanae* prompted us to study the effects of tungsten and selenium on *M. stadtmanae* growth.

Findings: Addition of 0.2 mg/L sodium tungstate to medium 322 yielded, 48 hours after inoculation, a growth rate equivalent to that obtained after 6 days with control culture as measured by methane monitoring and optical density measurement. Addition of 50 µg/mL sodium selenate had no effect on *M. stadtmanae* growth. Quantitative real-time PCRs targeting the *M. stadtmanae* 16S rRNA confirmed these data.

Conclusions: These data provide new information regarding the poorly known nutritional requirements of the human gut colonizing organisms *M. stadtmanae*. Adding sodium tungstate to basal medium may facilitate phenotypic characterization of this organism and additionally aid the isolation of new *Archaea* from complex host microbiota.

Keywords: Methanogenic *Archaea*, *Methanosphaera stadtmanae*, *Methanomassiliicoccus luminyensis*, Tungsten, Selenium

Findings

Methanosphaera stadtmanae is a spherical-shaped, non-motile archaeon initially isolated from human feces [1]. *M. stadtmanae* was the first human *Archaea* to be genome sequenced and analysis of the genome confirmed that *M. stadtmanae* belonged to *Methanobacteriales* [2]. PCR-based analyses further indicated that *M. stadtmanae*-specific sequences could be detected in stool specimen in up to 30% of individuals [3]. However, *M. stadtmanae* is a fastidious organism, with only one *M. stadtmanae* isolate reported and accordingly only one *M. stadtmanae* strain available in public collections. *M. stadtmanae* oxidizes hydrogen to reduce methanol into methane [1,2]. This metabolic trait has been already reported for *M. stadtmanae* [4], and more recently for members of the genus *Methanobacterium* (e.g. *M. veterum* and *M. lacus*;

[5,6]) within the order *Methanobacteriales*. We recently isolated *Methanomassiliicoccus luminyensis*, the first cultured representative of new order of methanoarchaea [7]. This archaeon exhibits a metabolic trait similar to that of *M. stadtmanae* by using hydrogen as electron donor and methanol as electron acceptor [7]. Unexpectedly, we observed that addition of tungstate-selenite to culture medium had been a key factor for successful isolation of *M. luminyensis* and that this archaeon indeed required tungstate-selenite as an essential element for growth. We therefore tested the hypothesis that the addition of sodium tungstate or sodium selenate or both to basal culture medium would also enhance the growth of *M. stadtmanae*.

M. stadtmanae DSMZ 3091^T (ATCC 43021^T) purchased from the German Collection of Microorganisms and Cell Cultures (DSMZ, Braunschweig, Germany) was grown on medium 322 (<http://www.dsmz.de>) incubated at 37°C in Hungate tubes (Dutscher, Issy-les-Moulineaux, France) under 2-bar pressure of a H₂/CO₂ (80–20) atmosphere. The inoculated medium (10% vol) was

* Correspondence: michel.drancourt@univmed.fr

¹Unité de Recherche sur les Maladies Infectieuses et Tropicales Emergentes UMR CNRS 6236 IDR 198, IFR48, Institut Méditerranée Infection, Aix-Marseille-Université, Marseille, France

Full list of author information is available at the end of the article

incubated at 37°C with shaking. On the exponential phase of this first culture, a second inoculation was performed by 10% vol. in the same basal medium modified or not by the addition of Na₂O₄W (0.2 mg/L) and/or Na₂O₄Se (50 µg/L) (Sigma, Saint-Quentin Fallavier, France). Non-inoculated media were used as negative controls and each experiment was repeated ten times.

Growth was assessed by optical microscope observation, parallel methane production measurement and measurement of the optical density of the medium. Methane production measurement used a GC-8A gas chromatograph (Shimadzu, Champs-sur-Marne, France) equipped with a thermal conductivity detector and a Chromosorb WAW 80/100 mesh SP100 column (Alltech, Carquefou, France). N₂ at a pressure of 100 kPa was used as the carrier gas. The detector and the injector temperatures were 200°C, and the column temperature was 150°C. H₂ consumption and CH₄ production were measured every 6 hours for 24 hours and then every 12 hours for 6 days. The optical density at 580 nm was measured by inserting Hungate tubes into the spectrophotometer (Varian Cary50; Agilent Technologies, Massy, France). Experiment was done in triplicate and average optical density value for the three replicates was calculated.

M. stadtmanae DNA extraction, quantification and sequencing were performed as previously described based on specific quantitative real-time PCR targeting 16S rRNA gene [3].

Negative controls (with and without tungstate and selenium) remained negative with no growth occurring after one-week incubation indicating that results herein reported did not merely result from carry-over of organisms. The exponential phase of *M. stadtmanae* growth cultured in medium 322 was reached at 6-day incubation. At this point microscopic observation disclosed organisms with morphology compatible with *M. stadtmanae* and no contaminant. Also, qPCR detected an equivalent of 3.22E + 12 ± 1.53E + 11 copies of 16S rRNA gene/mL (Table 1). Sequencing of 16S rRNA gene PCR products from all specimens yielded a sequence similarity of 99-100% with the reference *M. stadtmanae* DSM 3091 sequence.

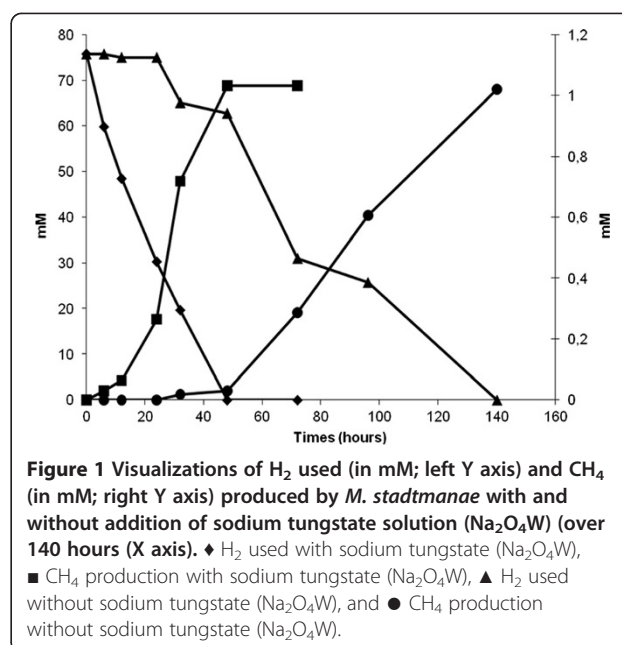


Figure 1 Visualizations of H₂ used (in mM; left Y axis) and CH₄ (in mM; right Y axis) produced by *M. stadtmanae* with and without addition of sodium tungstate solution (Na₂O₄W) over 140 hours (X axis). ♦ H₂ used with sodium tungstate (Na₂O₄W), ■ CH₄ production with sodium tungstate (Na₂O₄W), ▲ H₂ used without sodium tungstate (Na₂O₄W), and ● CH₄ production without sodium tungstate (Na₂O₄W).

The addition of sodium selenate alone has no effect on the growth curve of *M. stadtmanae*. However, the addition of sodium tungstate alone or in combination with sodium selenate shortened the lag period to 2 days post-inoculation with an equivalent 16S rRNA and *rpoB* genes copy number and with equivalent rates of methane production (Figure 1). In the absence of tungstate, *M. stadtmanae* exhibited a 30-hour log phase. Adding tungstate to the culture medium reduced the delay of this log-phase so that it took 47 hours instead of 72 hours to achieve a 0.35 optical density of the culture (Figure 2). These results correlated with the fact that *M. stadtmanae* genome encodes a formylmethanofuran dehydrogenase comprising of five sub-units (Genes IDs: 3855499-3855500-3855501-3855502-3855503), an enzyme found in methanogenic *Archaea*. In strict anaerobic micro-organisms, this enzyme catalyzes the reversible dehydrogenation of formylmethanofuran into CO₂ and methanofuran. The formylmethanofuran dehydrogenases are either molybdenum- or tungsten-iron-sulfur proteins. The tungsten is likely bound to the same

Table 1 *M. stadtmanae* 16S rDNA gene copy number after 48-hour culture with Na₂O₄W + Na₂O₄Se or only Na₂O₄W and a 6-day culture with no Na₂O₄W + Na₂O₄Se or only with Na₂O₄Se (Mean and standard deviation were calculated for 10 independent culture tests for each condition)

	48-hour culture			6-day culture	
	without Na ₂ O ₄ W + Na ₂ O ₄ Se	with Na ₂ O ₄ W + Na ₂ O ₄ Se	with Na ₂ O ₄ W	with Na ₂ O ₄ Se	without Na ₂ O ₄ W + Na ₂ O ₄ Se
Means	2.13E + 10	4.42E + 12	3.93E + 12	4.02E + 12	3.22E + 12
Standard deviation	5.56E + 09	1.84E + 11	3.67E + 11	2.23E + 11	1.53E + 11

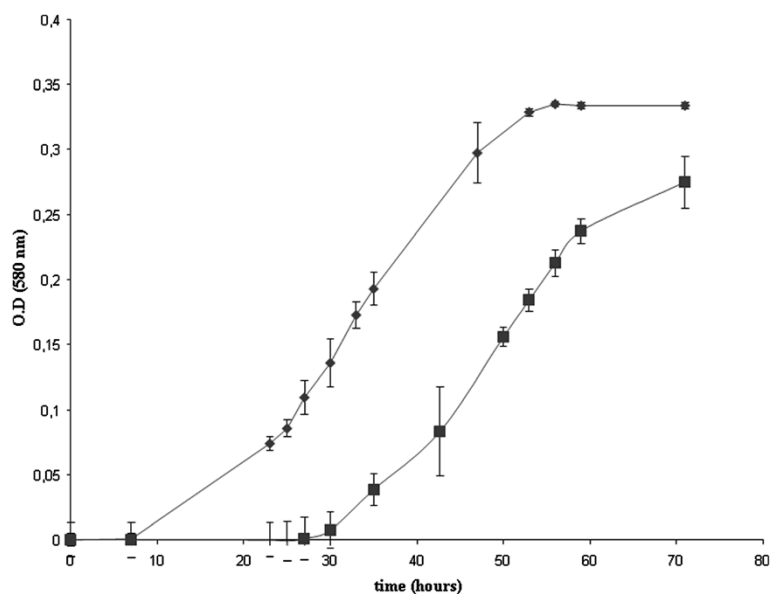


Figure 2 The effect of addition of selenite/tungstate solution on growth of *M. stadtmanae*. ◆ Growth of *M. stadtmanae* with tungstate ($\text{Na}_2\text{O}_4\text{W}$). ■ Growth of *M. stadtmanae* without tungstate ($\text{Na}_2\text{O}_4\text{W}$).

skeleton as the molybdenum in the so-called molybdopterin dinucleotide cofactor [8-10].

Previous reports described the requirement of tungsten for growing numerous methanogens including *Methanothermobacter wolfei* which has an obligate requirement for tungsten to maintain autotrophic growth, *Methanococcus vannielii* requiring tungsten as a cofactor for the enzyme formate dehydrogenase [11], *Methanogenium tatii* [12] and *Methanocorpusculum parvum* [13] also requiring tungsten for growth (Table 2). Selenium has also been reported as stimulatory and may be required for many methanogens, especially members of the genus *Methanococcus* as *Methanococcus vannielii* [11], *Methanococcus jannaschii* [14], *Methanococcus maripaludis* [15], *Methanococcus voltae* [16] and *Methanococcus thermolithotrophicus* [17] (Table 1). Requirement for selenium could have enzymatic basis, since it was reported that *M. vannielii* possesses a selenium-dependant formate

dehydrogenase [18]. Selenium was also reported as a component of both a hydrogenase [19] and tRNA [20].

In the absence of tungstate, *M. stadtmanae* exhibited a growth delay of 5–7 days which is long for testing *in vitro* susceptibility to antibiotics [22]. As we now observed that tungsten enhances the growth of two taxonomically unrelated methanogens, *M. stadtmanae* and *M. luminyensis*, we suggest that tungsten-containing media could be incorporated into the panel of media used for the isolation and culture of new methanogens from clinical and environmental specimens, and for testing their *in-vitro* susceptibility to antibiotics.

Methanogenic *Archaea* recently emerged as normal components of the human gastrointestinal and oral microbial ecosystems, where they could play important roles in health and diseases [23]. However, the isolation of such organisms requires long incubation times and strict anoxic atmosphere and is hampered by the incomplete knowledge of their nutritional requirements [23]. In fact, the result obtained in the present study may prompt further phenotypic characterization including extended antibiotic susceptibility testing [22] and even allowing isolation of new *Archaea* in order to assess understanding their contribution in the physiology of complex human microbiomes and their potential role in the course of infections.

Competing interests

The authors declare that they have no competing interests.

Author's contributions

BD, SK, MLF designed and performed analyses, BO, MD interpreted data and wrote the draft. All authors read and approved the final manuscript.

Table 2 Requirement of tungsten or/and selenium for growth of methanogens as reported in bibliography

Species	Tungsten	Selenium	References
<i>Methanothermobacter wolfei</i>	YES	NA	[21]
<i>Methanococcus vannielii</i>	YES	YES	[11,18]
<i>Methanogenium tatii</i>	YES	NA	[12]
<i>Methanocorpusculum parvum</i>	YES	NA	[13]
<i>Methanococcus jannaschii</i>	NA	YES	[14]
<i>Methanococcus maripaludis</i>	NA	YES	[15]
<i>Methanococcus voltae</i>	NA	YES	[16]
<i>Methanococcus thermolithotrophicus</i>	NA	YES	[17]

Author details

¹Unité de Recherche sur les Maladies Infectieuses et Tropicales Emergentes UMR CNRS 6236 IDR 198, IFR48, Institut Méditerranée Infection, Aix-Marseille-Université, Marseille, France. ²Laboratoire de Microbiologie IRD, UMR D180, Microbiologie et Biotechnologie des Environnements Chauds, Aix-Marseille-Université, ESIL, Marseille, France.

Received: 16 November 2011 Accepted: 25 April 2012
Published: 15 May 2012

References

1. Miller TL, Wolin MJ: *Methanosphaera stadtmaniae* gen. nov., sp. nov.: a species that forms methane by reducing methanol with hydrogen. *Arch Microbiol* 1985, **141**:116–122.
2. Fricke WF, Seedorf H, Henne A, Krueger M, Liesegang H, Hedderich R, et al: The genome sequence of *Methanosphaera stadtmanae* reveals why this human intestinal archaeon is restricted to methanol and H₂ for methane formation and ATP synthesis. *J Bacteriol* 2006, **188**:642–658.
3. Dridi B, Henry M, El Khéchine A, Raoult D, Drancourt M: High prevalence of *Methanobrevibacter smithii* and *Methanosphaera stadtmanae* detected in the human gut using an improved DNA detection protocol. *PLoS One* 2009, **4**:e7063.
4. Kendall M, Boone D: The Order *Methanosarcinales*. In *The Prokaryotes*. Edited by Dworkin M, Falkow S, Rosenberg E, Schleifer KH, Stackebrandt E. New York: Springer; 2006:244–256.
5. Krivushin KV, Shcherbakova VA, Petrovskaya LE, Rivkina EM: *Methanobacterium veterum* sp. nov., from ancient Siberian permafrost. *Int J Syst Evol Microbiol* 2010, **60**:455–459.
6. Borrel G, Joblin K, Guedon A, Colombet J, Tardy V, Lehours AC, Fonty G: *Methanobacterium lacus* sp. nov., a novel hydrogenotrophic methanogen from the deep cold sediment of a meromictic lake. *Int J Syst Evol Microbiol* 2011, Sep 2. [Epub ahead of print] PubMed PMID: 21890730.
7. Dridi B, Fardeau ML, Ollivier B, Raoult D, Drancourt M: *Methanomassiliicoccus luminyensis*, gen. nov., sp. nov., a novel methanogenic Archaea isolated from human feces. *Int J Syst Evol Microbiol* 2012, **62**:1902–1907. in press.
8. Karrasch M, Borner G, Enssle M, Thauer RK: The molybdoenzyme formylmethanofuran dehydrogenase from *Methanosarcina barkeri* contains a pterin cofactor. *Eur J Biochem* 1990, **194**:367–372.
9. Karrasch M, Borner G, Enssle M, Thauer RK: Formylmethanofuran dehydrogenase from methanogenic bacteria, a molybdoenzyme. *FEBS Lett* 1989, **253**:226–230.
10. Borner G, Karrasch M, Thauer RK: Molybdopterin adenine dinucleotide and molybdopterin hypoxanthine dinucleotide in formylmethanofuran dehydrogenase from *Methanobacterium thermoautotrophicum* (Marburg). *FEBS Lett* 1991, **290**:31–34.
11. Jones JB, Stadtman TC: *Methanococcus vannielii*: culture and effects of selenium and tungsten on growth. *J Bacteriol* 1977, **130**:1404–1406.
12. Zabel HP, König H, Winter J: Isolation and characterization of a new coccoid methanogen, *Methanogenium tatii* spec. nov. from a solfatara field on Mount Tatío. *Arch Microbiol* 1984, **137**:308–315.
13. Zellner G, Alten C, Stackebrandt E, Conway De Macario E, Winter J: Isolation and characterization of *Methanocorpusculum parvum* gen. nov., spec. nov., a new tungsten requiring, coccoid methanogen. *Arch Microbiol* 1987, **147**:13–20.
14. Jones WJ, Leigh JA, Mayer F, Woese CR, Wolfe RS: *Methanococcus jannaschii* sp. nov., an extremely thermophilic methanogen from a submarine hydrothermal vent. *Arch Microbiol* 1983, **136**:254–261.
15. Jones WJ, Paynter MJB, Gupta R: Characterization of *Methanococcus maripaludis* sp. nov., a new methanogen isolated from salt marsh sediment. *Arch Microbiol* 1983, **135**:91–97.
16. Berghöfer Y, Agha-Amiri K, Klein A: Selenium is involved in the negative regulation of the expression of selenium-free [NiFe] hydrogenases in *Methanococcus voltae*. *Mol Gen Genet* 1994, **242**:369–373.
17. Belay N, Sparling R, Daniels L: Relationship of formate to growth and methanogenesis by *Methanococcus thermolithotrophicus*. *Appl Environ Microbiol* 1986, **52**:1080–1085.
18. Jones JB, Stadtman TC: Selenium-dependent and selenium-independent formate dehydrogenases of *Methanococcus vannielii*. Separation of the two forms and characterization of the purified selenium-independent form. *J Biol Chem* 1981, **256**:656–663.
19. Yamazaki S: A selenium-containing hydrogenase from *Methanococcus vannielii*. Identification of the selenium moiety as a selenocysteine residue. *J Biol Chem* 1982, **257**:7926–7929.
20. Ching WM, Wittwer AJ, Tsai L, Stadtman TC: Distribution of two selenonucleosides among the selenium-containing tRNAs from *Methanococcus vannielii*. *Proc Natl Acad Sci U S A* 1984, **81**:57–60.
21. König H, Semmler R, Lerp C, Winter J: Evidence for the occurrence of autolytic enzymes in *Methanobacterium wolfei*. *Arch Microbiol* 1985, **141**:177–180.
22. Dridi B, Fardeau ML, Ollivier B, Raoult D, Drancourt M: The antimicrobial resistance pattern of cultured human methanogens reflects the unique phylogenetic position of archaea. *J Antimicrob Chemother* 2011, **66**:2038–2044.
23. Dridi B, Raoult D, Drancourt M: Archaea as emerging organisms in complex human microbiomes. *Anaerobe* 2011, **17**:56–63.

doi:10.1186/1756-0500-5-238

Cite this article as: Dridi et al.: Tungsten-enhanced growth of *Methanosphaera stadtmanae*. *BMC Research Notes* 2012 5:238.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

